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The Pathogenesis of Extraintestinal Manifestations: Implications for IBD Research, Diagnosis, and Therapy

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Abstract: This article reports on the sixth scientific workshop of the European Crohn's and Colitis Organisation [ECCO] on the pathogenesis of extraintestinal manifestations [EIMs] in inflammatory bowel disease [IBD]. This paper has been drafted by 15 ECCO members and 6 external experts [in rheumatology, dermatology, ophthalmology, and immunology] from 10 European countries and the USA. Within the workshop, contributors formed subgroups to address specific areas. Following a comprehensive literature search, the supporting text was finalized under the leadership of the heads of the working groups before being integrated by the group consensus leaders.

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The Pathogenesis of Extraintestinal Manifestations: Implications for IBD research, diagnosis and therapy.

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Short title: EIM pathogenesis in IBD

Non-standard abbreviations:

CD	Crohn's disease
IBD	Inflammatory bowel disease
TNF	Tumour necrosis factor
PSC	Primary sclerosing cholangitis

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MANUSCRIPT BODY

This article reports on the sixth scientific workshop of the European Crohn's and Colitis Organisation (ECCO) on the pathogenesis of extraintestinal manifestations (EIM) in inflammatory bowel disease (IBD). This paper has been drafted by 15 ECCO members and 6 external experts (rheumatology, dermatology, ophthalmology and immunology) from 10 European countries and the USA. Within the workshop contributors formed subgroups to address specific areas. Following a comprehensive literature search, the supporting text was finalised under the leadership of the heads of the working groups before being integrated by the group consensus leaders.

INTRODUCTION

Up to 50% of IBD patients experience at least one extraintestinal manifestation (EIM).¹ The pathogenic mechanisms of EIM are not clearly defined. Unravelling these pathways has the potential to enhance our understanding of the pathogenesis not only of EIM but also of IBD overall. Defining pathogenic pathways in EIM is challenging due to the lack of consistent criteria for diagnosis and difficulty in distinguishing drug-induced extraintestinal pathologies from EIM. Optimising treatment may also be problematic. For many EIMs commonly accepted definitions and high quality evidence supporting different treatment strategies are lacking.² Therefore, there is a great need for both basic science studies and clinical trials to understand pathogenesis and determine optimal treatment of EIM. The first ECCO European Evidence-based consensus on EIM in IBD provided an authoritative guideline for the clinical management of EIM.² The current article seeks to complement and extend the clinical guideline by identifying frontiers and open questions for clinical research.

DEFINITION

In order to standardize systematic inclusion of patients in scientific and clinical studies and align outcome measures to ensure clarity across the scientific literature, widely agreed upon definitions of the pathology being studied are critical. In order to provide a frame of

reference for scientific discourse the expert panel suggests the following *mechanistic* definition of what constitutes an EIM:

“An inflammatory pathology in a patient with IBD that is located outside the gut whose pathogenesis is either dependent on extension/translocation of immune responses from the intestine, or is an independent inflammatory event perpetuated by IBD or that shares a common environmental or genetic predisposition with IBD.”

A wide range of extraintestinal pathologies are associated with IBD, however not all of these would be considered to be true EIM according to the definition above. The panel proposes that current data supports pathologies listed in the first column of [Table 1](#) as true EIM, with other pathologies classified as associated auto-immune conditions or complications of IBD and its treatment. The distinction between these categories can be imprecise and overlap likely exists; it is probable that with future new data some pathologies will be re-classified. For the purpose of this review the panel focussed on true EIM as described by the definition above.

BASIC MECHANISMS OF EIM

Immunological mechanisms

The potentially diverse immune mechanisms that underlie EIM are poorly defined. We discuss two distinct theories that mechanistically link inflammation in the intestine and at other sites. First, EIM arise from an extension of antigen specific immune responses from the intestine to non-intestinal sites. Second, EIM are independent inflammatory events initiated or perpetuated by presence of IBD or by shared genetic or environmental risk factors in the host. These mechanisms are not mutually exclusive and may contribute to varying degrees in different EIM ([Figure 1](#)).

Extension of immune responses from the intestine

Ectopic expression of gut-specific chemokines and adhesion molecules:

Abnormal patterns of lymphocyte homing in IBD may contribute to EIM³. Expression of the vascular addressin MAdCAM-1 is normally restricted to intestinal tissue and, in the context

of specific chemokine signals, enables gut tropic T cells that express $\alpha 4\beta 7$ integrin to traffic selectively to the intestinal mucosa. Additional tropism for the small intestine comes from the chemokine CCL25 attracting lymphocytes expressing its receptor CCR9. Ectopic expression of both chemokines and adhesion molecules can occur in IBD⁴, and may facilitate trafficking of inflammatory T cells to extraintestinal sites. The best supporting evidence comes from IBD associated primary sclerosing cholangitis (PSC). Ectopic expression of both MAdCAM-1 and CCL25 has been demonstrated in the vascular endothelium of the portal tract.^{5, 6} One fifth of the infiltrating T cells co-express CCR9 and $\alpha 4\beta 7$, whereas the frequency of these cells is low in other forms of liver inflammation, indicating an important role for these molecules in recruitment of inflammatory lymphocytes in PSC.⁶ While attractive to propose ectopic expression of gut-associated addressins at extraintestinal sites as a logical mechanism for EIM, evidence that this occurs in organs other than liver is lacking. However, co-expression of $\alpha 4\beta 7$ with cutaneous leukocyte antigen (CLA), (implicated in homing to the skin), by some blood T cells from IBD patients⁷, may indicate that gut generated effector cells can acquire both gut and skin tropism.

T cell trafficking driven by non-specific adhesion molecules:

Upregulation of inflammation-associated adhesion molecules and chemokines that lack tissue restriction may also enable capture of effector cells facilitating their recruitment into non-intestinal sites. Gut leukocytes from IBD patients are able to bind to the synovial membrane, using a repertoire of adhesion molecules⁸, but mainly using endothelial vascular adhesion protein 1 (VAP-1)⁹. VAP-1 also plays a role in transmigration of lymphocytes across hepatic endothelium and its expression is upregulated by inflammation.^{10, 11} T cells from intestinal mucosa of IBD patients express chemokine receptors, such as CXCR3 and CCR5,^{12, 13} that may contribute to their ability to enter other tissues. Low-grade inflammation, injury or mechanical stress at extraintestinal sites (as implicated in the pathogenesis of spondyloarthritis (SpA)¹⁴ and pyoderma gangrenosum, where this phenomenon is termed pathergy) may nucleate the recruitment of gut-generated effector cells and further enhance the inflammatory process.

Microbial antigen translocation and/or cross-reactivity:

Models of EIM that invoke trafficking of gut effector T cells raise the question as to whether this process is dependent on antigen specific reactivation at non-intestinal sites and if so,

what the antigen may be. Antigens derived from the gut microbiota are believed to be key targets for intestinal effector T cells in IBD and transport of these antigens to the liver via the portal circulation may activate such cells localised here via $\alpha 4\beta 7$ -MAdCAM-1 interactions and other pathways. The presence of distinct gut microbiota in IBD patients with PSC¹⁵⁻¹⁷ may suggest specific bacterial antigens. At other sites, cells may be reactivated by cross-reactive components of the resident microbiota or host antigens. Molecular mimicry, in the form of peptide sequences common between enteric bacteria and host MHC molecules has been reported^{18, 19} although the pathologic significance of this is unclear. In mice, retina-specific T cells that cause uveitis require activation in the gut by a microbiota dependent signal, most likely a cross reactive bacterial antigen,²⁰ providing evidence for a direct link between gut microbiota, recognition of self-antigens and inflammation at a non-intestinal site. Indeed, leukocyte trafficking between the gut and the eye has been demonstrated in experimental models of autoimmune uveitis.²¹ However, the antigen specificity of T cells responsible for EIM in humans has never been defined.

Circulating antibodies:

Circulating antibodies could extend intestinal immune responses to additional sites and immune complex mediated inflammation has been proposed to contribute to certain EIM²². Autoantibodies reactive to colonic proteins have been identified in patients with IBD^{23, 24} and, using monoclonal antibodies, epitopes shared between human colon and tissues such as eyes, joints, skin and biliary epithelium have been identified.²⁵⁻²⁷ However, clear evidence of a causative role for antibodies or immune complexes in the pathogenesis of EIM in IBD patients is lacking.

EIM as independent inflammatory events

A shift in inflammatory tone favours the development of EIM:

An alternative explanation for EIM would see them as independent inflammatory events sharing common genetic²⁸ or environmental²⁹ risk factors with IBD. The presence of intestinal inflammation and/or microbial dysbiosis in individuals with IBD might further increase the risk of developing extraintestinal inflammation through modulation of inflammatory 'tone', impacting on immune functions at other sites. Key inflammatory mediators, including IL-6, TNF α , IFN γ and VEGF³⁰ are raised in serum of IBD patients, as is

bacterial LPS³¹ which may promote cytokine production via activation of immune cells at non-intestinal sites. Systemic effects, including increases in epithelial permeability³² and up-regulation of neutrophil extravasation ligands on vascular endothelium, may lower the threshold for immune activation at extraintestinal sites. IBD associated cytokines, such as IL-23, which is produced at high levels in CD and UC, can activate immune cells resident within the synovial membrane and drive spondyloarthritis.³³

Systemic changes in innate immune function:

Exposure of neutrophils to inflammatory cytokines or other signals can enhance their response upon subsequent activation, a phenomenon termed neutrophil priming.³⁴ Circulating neutrophils show morphological evidence of activation in IBD³⁵ and are primed to produce increased levels of TNF α and IL-1 β .³⁶ In contrast, recruitment of neutrophils to the skin and clearance of subcutaneous bacteria is reduced in patients with CD³⁷. Likewise, changes in circulating monocytes³⁸ and macrophages derived from blood monocytes³⁷ have been reported in IBD, with reduced inflammatory cytokine production in response to bacterial stimulation.

Altered haematopoiesis:

Changes in circulating immune cells observed in IBD are likely to reflect altered haematopoiesis in the bone marrow. In mouse models, haematopoiesis is influenced by both microbial products³⁹⁻⁴³ and the presence of intestinal inflammation.^{44, 45} In IBD, systemic inflammatory cytokines, increased permeability of the intestine to microbial products or changes in the composition and metabolic products of the microbiota, could all influence the generation of innate immune cells.^{46, 47}

Dysbiosis and gut microbiota

The long-established link between gut infections with enteric pathogens such as *Salmonella*, *Campylobacter*, *Yersinia* and *Shigella* and reactive arthritis is a clear indication that potential pathogenic pathways between microbiota in the gut and extraintestinal inflammation exist. Specific EIM are associated with gut dysbiosis: Patients with SpA have decreased faecal gut microbial diversity and increased abundance of *Ruminococcus gnavus* and genus *Dialister*, which positively correlated with disease activity.^{48, 49} Patients with psoriatic arthritis also

exhibit decreased faecal microbial diversity.⁵⁰ In addition, faecal *Saccharomyces cerevisiae* abundance is decreased in psoriasis compared to healthy controls.⁵¹

PSC has also been associated with decreased faecal microbial diversity.^{15, 52} One study has suggested that IBD patients have similar dysbiosis to IBD-PSC patients.¹⁷ However, a conflicting report⁵³ makes it difficult to judge whether the risk for developing PSC is driven by specific microbial factors.¹⁷ There is a paucity of studies determining gut dysbiosis in individuals with inflammatory eye disease, although one study demonstrated differences in gut microbiota between healthy individuals and those with age-related macular degeneration⁵⁴ and preliminary data suggested the existence of an intraocular microbiota.⁵⁵ There is a range of potential mechanisms by which gut microbiota drive the pathogenesis of EIM (**Box 1**).

1. Molecular mimicry: similarity between gut microbiota and non-microbial epitopes present at the extraintestinal site
2. Microbial communities in the extraintestinal site: similarities with pro-inflammatory gut microbiota could drive extraintestinal inflammation
3. Microbial translocation: Microbiota or their components are translocated from the gut to the extraintestinal site, (e.g. to the liver via the portal circulation)
4. Soluble microbial derived factors: e.g. LPS may be released into the circulation and promote inflammation at extraintestinal sites.
5. Disruption of gut barrier: Specific microbiota such as mucin degraders may disrupt the gut mucosal barrier, facilitating leakage of cellular or non-cellular factors into the circulation
6. Microbiota-derived metabolites: e.g. metabolism of bile acids and generation of short-chain fatty acids which could both alter immune signalling
7. Acquisition of deleterious microbiota in early life resulting in altered immune development, which in turn generates a persistent pro-inflammatory immune “tone”

The first four of these hypothesized mechanisms have been included in the discussion in the preceding section. However, gut microbiota may promote inflammation at extraintestinal sites through metabolic activities. *Ruminococcus* which is altered in patients with arthritis could initiate breach of the intestinal barrier through mucin degradation.⁵⁶ In rats gut

microbiota-dependent alterations in bile acid deconjugation are associated with altered bile acid profiles in extraintestinal sites including kidney, heart, plasma and liver, demonstrating that gut microbial metabolic functions have the potential to influence immune signalling at distant sites.⁵⁷ Short-chain fatty acids (SCFA) produced by many gut bacteria may have metabolic or immunomodulatory effects. In experimental autoimmune uveitis oral administration of SCFA attenuated uveitis severity and was associated with suppression of effector T cell induction.²¹ Furthermore, SCFA have a potential role in modulating T cell trafficking to extraintestinal sites. Finally it has been hypothesised that IBD linked dysbiosis may exert its pathogenic effect during immune development.⁵⁸ This is supported by animal models with bacterial colonisation of mice at age 3 weeks resulting in a persistent inflammatory tone, whereas colonisation at age 1 week did not.⁵⁹ Thus disruption of acquisition of gut microbiota early in life may generate persistent aberrant immune responses manifested in the gut or extraintestinally, or both. Indeed, factors that may influence the process of gut microbiota acquisition in early life such as breast feeding have been shown to be protective against the occurrence of ankylosing spondylitis (AS).⁶⁰

Open questions:

1. Are the gut microbiota pathogenic in EIMs (via any of the mechanisms mentioned in the text) or are EIMs independent of gut microbiota?
2. If microbiota play a role, what is the mechanism?
3. If EIMs are driven by microbiota are these the same or different from those involved in IBD pathogenesis?
4. Are microbial communities in other parts of the body involved in IBD pathogenesis?

Genetic basis of extraintestinal manifestations

Familial and epidemiological evidence:

There is an extensive overlap in genetic risk loci for both IBD and EIM, particularly AS.⁶¹ Association studies revealed a concordance in EIM present in 70% of parent-child pairs and in 84% of sibling pairs highlighting the role of genotype,⁶² (or early life environmental factors). In addition, the appearance of one EIM increases the probability of developing other EIM.^{1, 63} Further supporting the genetic underpinning of EIM, the CD risk gene NOD2, encoding a pattern recognition receptor, has also been associated with sacroiliitis,⁶⁴ and

uveitis.⁶⁵ Several HLA genes and HLA independent loci have been associated with the presence of EIM and a detailed description can be found in the Supplementary information. The genetic contribution to the pathogenesis of EIM and IBD comprises a combination of overlapping and independent loci, a situation which is consonant with the occurrence of EIM in individuals both with and without evidence of gut inflammation. However, whether the involved loci all contribute to pathology in an EIM-specific fashion, or whether there are genes that liberate inflammatory responses from restriction to specific body compartments and thus give rise to EIM in general, is not known.

Open questions:

1. Are the genes that predispose to specific EIM in IBD patients the same as the genes that predispose towards the EIM pathology in non-IBD patients?
2. Are there genes common to all EIM patients and distinct from non EIM IBD (immune mobility/ promiscuity factors)?
3. Do IBD patients with no EIM have protective factors i.e. they have the same genetic risk as EIM patients but have additional (genetic or environmental) protective factors?

Animal models of EIM

Animal models where inflammation is manifested at more than one anatomical site or bodily system (multifocal inflammation) provide experimental platforms to dissect pathogenic pathways of EIM and serve as tools to test potential therapies. However, only few models manifest multifocal inflammation, with colitis-arthritis models being the dominant phenotype available.

TNF^{ΔARE} mice carry a genetic deletion of TNF AU-rich elements (ARE), leading to overexpression of TNF.⁶⁶ The resulting phenotype is CD-like transmural and granulomatous chronic ileitis along with spondyloarthropathy-like sacroiliitis, Achilles tendon enthesitis and peripheral arthritis. Paradoxically (given the importance of innate immune responses in human IBD), in this model ileitis appeared to be dependent on the presence of mature T and/or B cells, as mice with TNF^{ΔARE} in combination with a RAG^{-/-} background developed only arthritis.⁶⁶ Furthermore, mice with intestinal epithelial cell-specific TNF ARE deletion develop ileitis but not EIM,⁶⁷ indicating that intestinal inflammation *per se* is not sufficient

for induction of arthritis, which is therefore presumably dependent on local TNF production in the joint. Ileitis is abrogated in germ-free $\text{TNF}^{\Delta\text{ARE}}$ ⁶⁸ and $\text{TNF}^{\Delta\text{ARE}}/\beta 7^{-/-}$ mice,⁶⁹ but the effects of such manipulations on joint inflammation have not been reported yet. Taken together, in $\text{TNF}^{\Delta\text{ARE}}$ mice, gut and joint inflammation likely represent independent phenomena mediated by a common pro-inflammatory factor.

HLA-B27 transgenic rats develop spondylarthritis and colitis, but also gastritis, psoriasis and epididymitis.⁷⁰ In the intestinal mucosa there is increased production of pro-inflammatory cytokines (IFN- γ , IL-2, IL-1 α , IL-1 β , TNF α , and MIP2) and in addition plasma concentrations of TNF α and IL-6 are raised. IL-23 and IL-17A may play important roles, in association with HLA-B27 misfolding in the ER and activation of the unfolded protein response, leading to downstream inflammation.^{71, 72} Interestingly, in this model both colitis and arthritis (but not dermatitis and epididymitis) are dependent upon the presence of microbiota.⁷³ The HLA-B27 model is consistent with a common genetic origin of multi-organ inflammation, but also emphasises the fact that some but not all EIM are dependent on microbiota. However, when interpreting data from germ-free models it is important to consider that conventionally reared mice are not only colonised with microbiota in the gut but also other organs such as skin, joints and eye which may also play a role in pathogenesis. More detailed experiments may be required to determine the contribution of extraintestinal microbiota communities in animal models of inflammation.

SKG mice that receive intraperitoneal injections of 1,3- β -glucan develop ileitis in association with enthesitis, arthritis, dactylitis, fasciitis, vertebral inflammation, and uveitis.⁷⁴ Treatment with anti-IL-23 mAbs or genetic deletion of the downstream cytokine IL-17A abrogate both ileitis and arthritis.⁷⁵ Time course expression studies identified intestinal mucosa as the source of elevated IL-23 production.⁷⁵ Nevertheless, immunological pathways of joint and gut inflammation in this model are not identical, as IL-22 neutralization reduced the severity of enthesitis but exacerbated ileitis in 1,3- β -glucan-treated SKG mice.

Animal models: Open questions

1. Could further animal models with intestinal inflammation and extraintestinal involvement (including sites other than joints) be developed?
2. Which common pathways between mucosal and extraintestinal inflammation are implicated in animal models where both occur?

3. What is the role of microbiota (including faecal transplant) in the development of inflammation in animal models?
4. Can animal models be used to elucidate the temporal relationship between intestinal disease and development of EIM?
5. How should animal models be used to investigate novel mechanisms and therapies such as neuroimmunomodulation?

Implications of the therapeutic effect of biologics and other treatments for EIM

Emerging data for the efficacy of biologics for the treatment of EIM may serve to expose underlying pathogenic mechanisms. Most evidence is available for anti-TNF α with good response rates for cutaneous manifestations, arthritis and ocular EIM. This has implicated TNF α -dependent mechanisms in EIM pathophysiology.^{69-71, 76} However, anti-TNF α drugs are increasingly recognised to cause drug-induced skin lesions, contributing to the burden of skin disease in IBD.^{77, 78} The pathogenesis of these lesions remains unclear; blocking TNF α may result in an imbalance of cytokines (for example, increased IFN α release, which can cause psoriasis)⁷⁹⁻⁸¹ and TNF α inhibition may lead to a reduced accumulation of Th1 and Th17 cells at the site of inflammation, but trigger a compensatory expansion at other locations.⁸² Female gender and family history of inflammatory skin disorders were identified as risk factors which may also indicate a possible genetic predisposition for anti-TNF α induced skin lesions.⁸³

The gut selective mechanism of the integrin $\alpha 4\beta 7$ antibody vedolizumab should restrict its activity to the gut, since its counterpart MAdCAM1 is not expressed in the human skin.⁸⁴ The contribution of vedolizumab trials to understanding of EIM pathogenesis is complicated, since the evidence of its effect on EIM appears to be conflicting: One case series did not show any positive effect,⁸⁵ while a recent analysis from France suggested positive effects on EIM in most cases, but also revealed new onset of arthritis and paradoxical skin lesions.⁸⁶ The pathogenic mechanisms behind these observations remain elusive.⁸⁵ It may be speculated that a compensatory expansion of T cells at locations other than the gut could explain this phenomenon (similar to anti-TNF α induced lesions). On the other hand, a beneficial effect of vedolizumab on disease activity of EIM could occur if lymphocytes require the $\alpha 4\beta 7$ - MAdCAM1 interaction to gain access to the gut where they are activated,

followed by non- $\alpha 4\beta 7$ -dependent entry to extraintestinal sites. There is also evidence in animal models that some regulatory T cells require $\alpha 4\beta 7$ -dependent entry into the gut to be educated before expressing their function elsewhere; vedolizumab could theoretically interfere with this.^{87, 88} An alternative hypothesis is that $\alpha 4\beta 7$ is directly involved in homing to extraintestinal sites as outlined above. It remains likely that vedolizumab has the capacity to illuminate pathogenic pathways in EIM.

Data on other biological agents are limited. So far, no trial has been published evaluating the anti-IL12/23 antibody ustekinumab in the management of EIM. Case series suggest it has efficacy in the treatment of anti-TNF α induced skin lesions,^{89, 90} however, development of pustular psoriasis has been described.⁹¹ Whether ustekinumab is effective in the treatment of non-drug induced EIM has yet to be determined. In contrast to anti-IL12/23 and despite the pathogenic role of Th17 cells in the development of colitis, trials with anti-IL-17A have failed in IBD with even higher adverse rates than placebo.⁹² Moreover, in contrast to its efficacy in other inflammatory disorders, anti-IL-17A can even exacerbate IBD activity,⁹³ which highlights a distinct involvement of the IL-17A pathway in these entities. No data on JAK inhibitors is available so far, but upregulation of STAT3 in erythema nodosum and pyoderma gangrenosum⁸⁴ makes a response to JAK inhibitors reasonable to predict and sheds light on the possible involvement of the JAK-STAT pathway in cutaneous EIM.

Taken together, it is important that clinical trials and observational studies of biologic agents are designed to optimise the capture of data on effects on inflammation in other systems than the disease defined in the primary outcome.

Open questions:

1. How does vedolizumab affect EIM? Does it have the same effect on all EIM?
2. What is the implication of the overexpression of STAT for the prospect of using JAK-inhibitors for treating PG and EN?
3. How will EIM respond to IL12/23 treatment?

CLINICAL RESEARCH

Despite the presence of a wide range of EIM associated with IBD, standardized criteria for diagnosis, documentation or monitoring are lacking. Thus far only one randomized

controlled trial including IBD patients with EIM has been conducted.⁹⁴ Here we discuss the currently available paradigms and tools for clinical research in three of them: skin, joint and eye EIMs.

Diagnosis and monitoring of EIM

Because the diagnostic and monitoring tools for EIM have been developed within the organ-based specialties, this section is presented according to an organ-based structure.

Clinical criteria, indexes and scales

Joint manifestations:

IBD-associated joint symptoms may be subdivided into inflammatory and non-inflammatory joint pain, (arthritis and arthralgia respectively).^{95, 96} Inflammatory arthropathies in IBD are the most common EIM and belong to the spondyloarthritis (SpA) group with a prevalence of 20-50% for axial inflammation,⁹⁷⁻⁹⁹ and 5-20% for peripheral arthritis.^{100, 101} The Assessment of SpondyloArthritis International Society (ASAS) developed classification criteria for both inflammatory axial and peripheral joint disease. These criteria are the current standard for clinical trials research and have good performance as tested against the rheumatologist's diagnosis ([Supplementary Figures 1 and 2](#)).¹⁰²⁻¹⁰⁴ However, limited data evaluate ASAS criteria specifically in IBD patients. In IBD patients with inflammatory back pain ASAS criteria have an equivalent sensitivity but lower specificity compared to non-IBD patients.¹⁰⁵ This lower specificity may be due to the inclusion of IBD as one of the ASAS criteria of axial spondyloarthritis. Alternative classification tools such as the Amor classification¹⁰⁶ and the European Spondyloarthropathy Study Group (ESSG) criteria¹⁰⁷ also include IBD as a criterion, whereas the older Modified New York classification do not.¹⁰⁸ Nevertheless, in order to ensure applicability of research data to clinical practice it is advantageous that the definition of patient groups in clinical trials and research is consistent with that used rheumatology (i.e. ASAS criteria). Therefore, validation of these currently used tools in IBD patients should be carried out.

Monitoring tools for determining response to treatment and disease outcomes have also been developed by ASAS. The current gold standard tool for axial spondyloarthritis is the Ankylosing Spondylitis Disease Activity Score (ASDAS),¹⁰⁹ providing both a measurement of disease activity that may be followed over time as well as cut-offs to allow grouping of patients into different disease activity states ([Supplementary Figure 3](#)). ASDAS includes back

pain as one of the criteria. Hence, it is not well-adapted for use in the 5-20% of IBD patients with peripheral arthritis. In response to the lack of validated outcome measures in peripheral spondyloarthritis the authors of one randomized controlled trial of adalimumab in patients with non-psoriatic peripheral spondyloarthritis developed a new outcome measure, the Peripheral SpA Response Criteria (PSpARC40) measured after 12 weeks of treatment.¹¹⁰ However, this outcome measure has not been widely applied and there is a need to validate the use of these tools in patients with multifocal inflammation.

Eye manifestations:

The most common eye EIMs are episcleritis and anterior uveitis. Scleritis and posterior or intermediate uveitis are rarer, but pose a greater potential risk to sight. **Supplementary Figure 4** summarizes some of the more common types of inflammatory eye disease as well as some of the ocular complications of IBD and its treatment. Episcleritis is usually treated topically with corticosteroids or non-steroidals. Uveitis may pose a greater diagnostic and therapeutic challenge. The SUN (Standardisation of Uveitis Nomenclature) classification is internationally acknowledged and as such research and clinical trials in uveitis in IBD patients should follow this system (**Supplementary tables 2-7**).^{111, 112} SUN classification may be used both for diagnosis and classification of uveitis at presentation as well as for monitoring disease progression. However, it is relevant to consider that the SUN classification may have limitations especially for judging significance of the outcome of clinical interventions. The FDA defines a significant clinical response as 2-step change in parameters of the SUN classification, but many successful therapies do not meet the required 2 step improvement (especially in vitreous haze). Furthermore, the SUN classification describes anterior chamber cells as in unequal steps (0, +0.5, +1, +2 and +3) estimated subjectively by the consulting ophthalmologist which is therefore not optimal for quantitative research.

Skin manifestations:

Cutaneous manifestations are common in IBD patients¹¹³ and include ectopic cutaneous IBD in addition to the other categories of pathologies as set out in **Table 1**. The diagnosis of cutaneous manifestations is principally based on clinical examination of the patient due to the inherently accessible nature of the skin. In atypical cases a skin biopsy is helpful.¹¹⁴ In skin disorders, such as psoriasis and eczema specific indexes to objectively measure skin disease extent and activity have been developed (e.g. the Psoriasis Area Severity Index

(PASI)¹¹⁵⁻¹¹⁷ and the Eczema Area and Severity Index (EASI)).¹¹⁸ However similar standardised assessment techniques for cutaneous EIM of IBD such as EN and PG are lacking. The only randomised controlled trial of therapy for an EIM in IBD patients (infliximab for PG) employed a primary endpoint of clinical improvement at week 2, as determined by the clinician and patient's global assessment of reduction in ulcer size and depth and the degree of undermining of the ulcer edge.⁹⁴ Infliximab was shown to be superior to placebo, particularly in patients with disease duration ≤ 3 months. Standardisation of assessments methods such as that employed in this trial will enhance reproducibility in clinical research as well as facilitating meta-analysis of EIM research.

In summary, current tools for the diagnosis of EIM have for the most part been developed in patients with unifocal inflammation. Studies to validate the use of these tools in patients with multifocal inflammation, including IBD patients are needed. Even better would be a system of diagnosis and monitoring that reflects common pathogenic mechanisms which could then be applied to diseases generated by that common mechanism but manifesting in diverse clinical phenotypes.

Open questions:

1. Are tools for monitoring of unifocal inflammation valid for use in patients with multifocal inflammation?
2. If one of the criteria in an algorithm for diagnosing inflammatory pathology at an extraintestinal site is that the patient has IBD, will such an algorithm provide adequate diagnostic discrimination when applied to a population of IBD patients?
3. Is a single multidimensional scale for diagnosis and monitoring of inflammation at multiple sites possible? Is it desirable?

Biomarkers

There are no specific biomarkers for EIM activity in IBD with acute phase proteins ESR and CRP, leucocytosis, thrombocytosis and anaemia being non-specific and in addition ESR and CRP having low sensitivity being elevated in only 40-50% of patients with axial SpA. Conversely faecal calprotectin is only validated in the diagnosis and monitoring of gut inflammation and does not reflect disease activity at other sites.

Genetic markers for SpA:

Genetic factors may be utilised as biomarkers in the diagnosis of inflammatory pathology^{119, 120}. Combining clinical factors with genetic data has been shown to be superior in predicting the development of EIMs compared to either alone. HLA-B27 positive IBD patients are at increased risk for developing AS.² Apart from HLA-B27, over 41 genes have been identified predisposing to AS.^{121, 122} However, most of these have not been associated with increased risk for extra-articular inflammation. Currently there are neither reliable genetic biomarkers for peripheral SpA,^{123 119, 124} nor for cutaneous or ocular EIM.

Imaging biomarkers for spondyloarthritis (SpA):

Traditional X-rays are of value in diagnosing axial SpA but only demonstrate changes in advanced cases. MRI usually demonstrates the first radiological changes in axial SpA and is - despite moderate sensitivity and specificity¹²⁵ - the imaging test of choice for detection of early disease¹²⁶⁻¹²⁸ as well as the best objective technique to assess inflammatory disease activity.¹²⁹⁻¹³⁴ This assessment has been standardised with the use of the Bath Ankylosing Spondylitis Radiology Index (BASRI).¹³⁵ In peripheral arthritis which is generally non-erosive, joint radiography is usually normal so ultrasonography is often employed to confirm the diagnosis. In addition, there is no evidence to confirm or refute the assumption that radiological findings in inflammatory arthropathy differ between patients with patients with only arthritis and those with inflammation also at distal sites.

Antimicrobial antibodies:

IBD is associated with the presence of antibodies to a variety of microorganisms such as anti-Saccharomyces cerevesiae antibodies (ASCA), antineutrophil cytoplasmic antibodies (ANCA), anti-I2 (associated with anti-Pseudomonas activity), anti-Escherichia coli outer membrane porin C (anti-OmpC) and anti-flagellin antibodies (anti-CBir1). Subclinical intestinal inflammation has been reported to be present in a significant proportion of patients with radiographic axial SpA.^{136, 137} The data on the presence of these antimicrobial antibodies in patients with both IBD and SpA are inconsistent and mostly relate to axial SpA. Anti-I2 antibodies have been associated with the combination of AS and intestinal inflammation¹³⁷ as have antibodies against ASCA, anti-OmpC and anti-CBir1.¹³⁸

Open questions:

1. Should patients with presenting with inflammatory pathologies be screened for multifocal inflammation?
2. Which biomarkers would be most appropriate for screening and in which populations?
3. Would biomarkers be useful to guide therapeutic decisions even in patients with unifocal inflammation (to reveal underlying mechanisms)?

Predictors and treatment of EIM

Predictors of EIM

The identification of patients at risk of EIM is desirable as this raises the possibility not only of treatment initiation prior to permanent tissue destruction, but even the potential for disease prevention. Moreover, patients in whom a propensity to develop inflammatory disease has already declared itself in one system may provide a unique opportunity for targeted screening in order to detect inflammation at distant anatomical sites. Several studies have investigated factors influencing the risk of developing EIM but with inconsistent results. This is likely caused by differences across studies regarding definitions and assessment of EIM as well as patient populations since only very few population-based studies exist. Furthermore, the occurrence and risk factors for EIM may also vary geographically.¹³⁹⁻¹⁴¹

On the simplest level, demographic and clinical factors may be used to detect risk. For example, female sex,^{29, 63, 113, 142-146} CD rather than UC,^{113, 142, 143, 147, 148} increasing age,^{29, 143, 149} long disease duration,^{142, 143} colonic location in CD,^{100, 143} extensive UC compared to proctitis,^{142, 147} indicators of severe disease including need for steroids,¹⁴⁶ azathioprine,¹⁴⁶ biological therapy²⁹ or surgery,^{100, 144, 148} and smoking both in CD^{29, 148} and UC^{29, 150} have all been associated with an increased risk for EIM. However, these associations are not reported consistently and are not replicated in all population-based studies¹⁴⁷⁻¹⁴⁹ and as such, this approach may have limited applicability in clinical practice. Genetic factors play an important role in determining the presence of EIM,^{119, 120} especially genes in the HLA region on chromosome 6 as described above.¹⁵¹⁻¹⁵³ Combining clinical factors with genetic data has been shown to be superior to predict the development of EIM compared to either alone.^{119,}

¹²⁴ Furthermore, specific features of the clinical presentation may alert the clinician to the potential for future EIM. For example, IBD is in the differential of any patient with ocular inflammation, especially in the “typical” constellation of bilateral anterior/intermediate granulomatous uveitis. Conversely, it is wise to monitor liver function tests especially in the IBD patient presenting with the clinical picture of mild, extensive colitis with rectal sparing and backwash ileitis, often associated with PSC.

Screening for IBD in patients with AS has been studied with some success, although the low rate of development of IBD in this group made the usefulness of screening somewhat questionable.¹⁵⁴ EIM are often tested for based on clinical suspicion however, screening for secondary diagnoses in patients with inflammatory pathologies has not yet proved a fruitful strategy.

Open questions:

1. Can accurate predictors of EIM be developed?
2. Once predictors are available: Can early intervention alter the future development of EIM?

Treatment

A recent systematic review by Peyrin-Biroulet *et al.* based on nine interventional studies, seven open label studies and thirteen non-interventional studies found a good clinical efficacy of adalimumab and infliximab for the treatment of musculoskeletal, cutaneous, and ocular manifestations, and some beneficial effect in metabolic bone disease and haematological or vascular EIM in IBD patients.¹⁵⁵ In contrast, no or limited efficacy of other biologic drugs including certolizumab pegol, golimumab, vedolizumab or natalizumab was identified. In this review however different ranges of pathology get grouped together, which may obscure the therapeutic effect for specific types of EIM that share a common mechanism.

Paradoxical, drug-induced EIM are well documented and hint at the complex effects that interference with immune function may have. This complexity is potentially compounded in patients with multifocal inflammation. The effect of vedolizumab, (which blocks $\alpha 4\beta 7$ dependent migration of lymphocytes into the gut), on EIM has been difficult to predict, as

discussed above. Potentially vedolizumab may have no effect on extraintestinal inflammation due to its gut-selective nature; alternatively if lymphocytes causing extraintestinal inflammation require activation in the gut before migration to the distant site, then vedolizumab would be predicted to improve EIM. Lastly, if prevention of migration to the gut resulted in accumulation of lymphocytes at extraintestinal sites, then vedolizumab could cause exacerbation of EIM. Of course, it may be that each of these mechanisms is present in different patients. Another treatment strategy that may be examined in the future is combination therapy with biologics with different molecular targets, for example, combined anti-integrin/anti-TNF α therapy for IBD patients with EIM has shown some efficacy.¹⁵⁶

It has been hypothesized that the extent of inflammation (for example the size of ulcerations in PG) may determine optimal drug dosing, with larger ulcers requiring higher doses of the drug.⁹⁴ However no dose-response studies and no RCTs have been presented in IBD-EIM patients during the induction phase of anti-TNF α treatment to determine optimal trough levels.¹⁵⁷ The concept of relating drug dose to total inflammatory burden has instinctive validity and could be potentially of great relevance to patients with EIM. However, this concept remains speculative at present and requires validation in clinical trials.¹⁵⁸

Open questions:

1. Is there a dose-response relationship between anti-TNF α therapy and EIM treatment response?
2. Are all anti-TNF α antibodies equally effective for the treatment of EIM?
3. Is there an additive effect of combined immunosuppression in IBD patients with EIM?
4. Are optimal anti-TNF α trough levels for IBD patients with EIM different from those for IBD patients without EIM?

Treat to Target and Patient Reported Outcome Measures in EIM

A “treat to target” strategy has been developed in many areas of medicine, where treatment outcomes are defined by specific objective endpoints. The concept driving this strategy is that traditional outcome measures fail to reflect subclinical, yet active disease, permitting the accumulation of tissue damage over time. With a treat-to-target strategy therapy is

intensified until the relevant evidence-based treatment target is in the desired range, which is associated with a reduction in end-organ destruction. For example in rheumatoid arthritis and scores such as the Disease Activity Score Calculator for Rheumatoid Arthritis (DAS-28) have been established.¹⁵⁹ This approach has also been used successfully in endocrinology especially in diabetes management.¹⁶⁰⁻¹⁶² Ongoing studies are developing this strategy in IBD.¹⁶³ Whether the same treatment targets developed for unifocal inflammation, can be applied (individually or perhaps in combination) in IBD-EIM or whether different targets should be developed, is unclear.

Another current advancement in the care of patients with chronic conditions is the development of Patient Reported Outcome Measures (PROM), which may themselves function as a treatment target. PROM are defined by the FDA as “any report of the status of a patient’s health condition that comes directly from the patient, without interpretation of the patient’s response by a clinician or anyone else.”¹⁶⁴ PROM may be disease specific such as the Inflammatory Bowel Disease Questionnaire (IBDQ-32),¹⁶⁵ the Inflammatory Bowel Disease Quality of Life Questionnaire (IBDQOL)¹⁶⁶ or the Work Productivity and Activity Impairment: Crohn’s Disease (WPAI: CD).¹⁶⁷ The use of PROM designed for the assessment of unifocal inflammation in patients with multifocal inflammation presents obvious drawbacks, potentially missing significant aspects of the patient’s experience. However, non-disease specific instruments have been developed such as the Short Form Health Survey¹⁶⁸ and the EQ-5D,¹⁶⁹ which may be more applicable in multifocal inflammation.

Open questions:

1. What are appropriate treatment targets for patients with multifocal inflammation?
2. Can established treatment targets from patients with joint, skin or eye disease be employed for patients with EIM in IBD?
3. Would there be a difference in how PROM and treat to target strategies function in patients with EIM activity that is synchronous with the IBD activity compared with patients with asynchronous disease activity?

CONCLUSION

Determining the mechanisms that cause inflammation to manifest unifocally or multifocally in different patients remains an enticing conundrum in immunology. Solving this conundrum may illuminate novel mechanisms and reveal a broader range of therapeutic targets. In the

context of the availability of a greater number of drugs targeted toward this broadening range of molecular targets, the previous organ-based approach to inflammatory disease may be inadequate. A holistic approach to the diagnosis and monitoring of inflammatory disease will allow a personalised therapeutic strategy. New tools for monitoring multifocal inflammation are needed in order to better capture the experience of the patient. This holistic approach to inflammatory disease requires greater cooperation between specialities and across research disciplines.

Figures

Figure 1. See separate document

Figure legend

Figure 1 Potential mechanisms of EIM

I Extension of immune responses from the intestine

- A. Ectopic expression of adhesion molecules and chemokines *e.g. ectopic expression of MAdCAM-1 and CCL25 in the vascular endothelium of the portal tract*
- B. T cell trafficking driven by non-specific adhesion molecules *e.g. α 4 β 7-independent binding of leukocytes to the synovial membrane using a repertoire of adhesion molecules. Non-specific interactions may be initiated after low-grade inflammation, injury or mechanical stress*
- C. Microbial antigen translocation *e.g. via portal tracts*
- D. Microbial antigen cross reactivity *e.g. molecular mimicry between enteric bacteria and host MHC molecules*
- E. Circulating antibodies *that may bind epitopes shared between human colon and extraintestinal tissues*

II EIM as independent inflammatory events

- F. Shift in inflammatory tone *driven by genetic, environmental or microbial factors or by systemic increase in key inflammatory mediators*
- G. Systemic changes in innate immune function *e.g. neutrophil priming*
- H. Altered haematopoiesis *driven by microbial products, intestinal inflammation, systemic inflammatory cytokines, increased gut permeability, changes in the composition or metabolic products of the microbiota*
- I. Gut microbiota drives distant inflammation *via microbial products such as LPS, through changes in gut permeability, microbiota-derived metabolites*

Tables

Table 1. Suggested categorisation of extraintestinal conditions that occur in IBD patients, (list of extraintestinal conditions associated with IBD adapted from Harbord *et al.*²).

System	A. Extraintestinal manifestations (multifocal inflammation)	B. Complications of IBD and its treatment	C. Associated conditions with uncertain mechanism
Joints and bones	Spondyloarthritis	Metabolic bone disease/ osteoporosis - (drug or nutritionally induced)	Non-inflammatory arthralgia
Eye	Uveitis Episcleritis Scleritis	Drug induced cataracts and other drug-induced and nutritional eye disease (see supplementary figure 4)	
Oral, aural and nasal	Oral CD Orofacial granulomatosis Metastatic CD		Sensorineural hearing loss
Skin	Erythema nodosum Pyoderma gangrenosum Sweet syndrome Metastatic CD	Drug-induced skin disease (e.g. anti-TNF induced psoriasis, DILE) Drug-induced skin cancer Drug hypersensitivity	Vitiligo Psoriasis Eczema Epidermolysis bullosa acquisita Cutaneous polyarteritis nodosa Hidradenitis

			suppurativa
Urogenital	Metastatic CD	Nephrolithiasis Amyloidosis Drug-induced tubulo-interstitial nephritis	
Hepato-pancreato-biliary	PSC	Portal vein thrombosis Hepatic amyloidosis DILI Drug-induced pancreatitis	Autoimmune hepatitis Granulomatous hepatitis Autoimmune pancreatitis
Neurological		Peripheral neuropathy (drug or nutritionally induced) Venous sinus thrombosis Stroke	Central demyelination
Cardiovascular		Ischaemic heart disease Cerebrovascular accident Mesenteric ischaemia	
Pulmonary		Drug-induced lung fibrosis	Inflammatory bronchial and parenchymal lung disease including asthma, bronchiectasis and interstitial pneumonias
Coagulopathy		Venous thromboembolism	
Endocrine		Drug-induced Cushing's and Addison syndromes Drug-induced diabetes	Type 1 diabetes Autoimmune thyroid disease

Infection		Infections including systemic and local secondary to immunosuppression Septic complications of IBD or surgery	
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IBD: Inflammatory bowel disease, CD: Crohn's disease, DILE: Drug-induced lupus erythematosus, PSC: primary sclerosing cholangitis, DILI: Drug-induced liver injury

A. For several conditions there is evidence for a mechanistic link between two pathologies, as described by the definition put forward in this paper of a "true" extraintestinal manifestation (EIM) of IBD. We would propose that these conditions may also be considered multifocal inflammation.

B. Other conditions that occur in IBD patients are complications of the disease or its surgical or pharmacological management.

C. Several conditions occur more commonly in IBD patients but there is lack of evidence to categorise these as either complications or directly link them mechanistically to IBD. It is likely that as pathogenic mechanisms are better understood it may be possible to re-classify some of these conditions as "true" EIM/ multifocal inflammation.

REFERENCES

1. Vavricka SR, Rogler G, Gantenbein C, et al. Chronological Order of Appearance of Extraintestinal Manifestations Relative to the Time of IBD Diagnosis in the Swiss Inflammatory Bowel Disease Cohort. *Inflamm Bowel Dis* 2015;21:1794-800.
2. Harbord M, Annese V, Vavricka SR, et al. The First European Evidence-based Consensus on Extra-intestinal Manifestations in Inflammatory Bowel Disease. *J Crohns Colitis* 2016;10:239-54.
3. Adams DH, Eksteen B. Aberrant homing of mucosal T cells and extra-intestinal manifestations of inflammatory bowel disease. *Nat Rev Immunol* 2006;6:244-51.
4. Trivedi PJ, Bruns T, Ward S, et al. Intestinal CCL25 expression is increased in colitis and correlates with inflammatory activity. *J Autoimmun* 2016;68:98-104.
5. Grant AJ, Lalor PF, Hubscher SG, et al. MAdCAM-1 expressed in chronic inflammatory liver disease supports mucosal lymphocyte adhesion to hepatic endothelium (MAdCAM-1 in chronic inflammatory liver disease). *Hepatology* 2001;33:1065-72.
6. Eksteen B, Grant AJ, Miles A, et al. Hepatic endothelial CCL25 mediates the recruitment of CCR9+ gut-homing lymphocytes to the liver in primary sclerosing cholangitis. *J Exp Med* 2004;200:1511.
7. Mann ER, McCarthy NE, Peake ST, et al. Skin- and gut-homing molecules on human circulating gammadelta T cells and their dysregulation in inflammatory bowel disease. *Clin Exp Immunol* 2012;170:122-30.
8. Salmi M, Andrew DP, Butcher EC, et al. Dual binding capacity of mucosal immunoblasts to mucosal and synovial endothelium in humans: dissection of the molecular mechanisms. *J Exp Med* 1995;181:137-49.
9. Salmi M, Jalkanen S. Human leukocyte subpopulations from inflamed gut bind to joint vasculature using distinct sets of adhesion molecules. *The Journal of Immunology* 2001;166:4650.
10. McNab G, Reeves JL, Salmi M, et al. Vascular adhesion protein 1 mediates binding of T cells to human hepatic endothelium. *Gastroenterology* 1996;110:522-8.
11. Lalor PF, Edwards S, McNab G, et al. Vascular adhesion protein-1 mediates adhesion and transmigration of lymphocytes on human hepatic endothelial cells. *J Immunol* 2002;169:983-92.
12. Agace WW, Roberts AI, Wu L, et al. Human intestinal lamina propria and intraepithelial lymphocytes express receptors specific for chemokines induced by inflammation. *Eur J Immunol* 2000;30:819-26.
13. Yuan YH, ten Hove T, The FO, et al. Chemokine receptor CXCR3 expression in inflammatory bowel disease. *Inflamm Bowel Dis* 2001;7:281-6.
14. Jacques P, McGonagle D. The role of mechanical stress in the pathogenesis of spondyloarthritis and how to combat it. *Best Pract Res Clin Rheumatol* 2014;28:703-10.
15. Kummen M, Holm K, Anmarkrud JA, et al. The gut microbial profile in patients with primary sclerosing cholangitis is distinct from patients with ulcerative colitis without biliary disease and healthy controls. *Gut* 2017;66:611-619.
16. Torres J, Bao X, Goel A, et al. The features of mucosa-associated microbiota in primary sclerosing cholangitis. *Aliment Pharmacol Ther* 2016;43:790-801.
17. Kevans D, Tyler AD, Holm K, et al. Characterization of Intestinal Microbiota in Ulcerative Colitis Patients with and without Primary Sclerosing Cholangitis. *J Crohns Colitis* 2016;10:330-7.
18. Scofield RH, Kurien B, Gross T, et al. HLA-B27 binding of peptide from its own sequence and similar peptides from bacteria: implications for spondyloarthropathies. *Lancet* 1995;345:1542-4.
19. Ramos M, Alvarez I, Sesma L, et al. Molecular mimicry of an HLA-B27-derived ligand of arthritis-linked subtypes with chlamydial proteins. *J Biol Chem* 2002;277:37573-81.
20. Horai R, Zarate-Blades CR, Dillenburg-Pilla P, et al. Microbiota-Dependent Activation of an Autoreactive T Cell Receptor Provokes Autoimmunity in an Immunologically Privileged Site. *Immunity* 2015;43:343-53.
21. Nakamura YK, Janowitz C, Metea C, et al. Short chain fatty acids ameliorate immune-mediated uveitis partially by altering migration of lymphocytes from the intestine. *Sci Rep* 2017;7:11745.
22. Bodenheimer HC, Jr., LaRusso NF, Thayer WR, Jr., et al. Elevated circulating immune complexes in primary sclerosing cholangitis. *Hepatology* 1983;3:150-4.
23. Biancone L, Mandal A, Yang H, et al. Production of immunoglobulin G and G1 antibodies to cytoskeletal protein by lamina propria cells in ulcerative colitis. *Gastroenterology* 1995;109:3-12.

24. Geng X, Biancone L, Dai HH, et al. Tropomyosin isoforms in intestinal mucosa: production of autoantibodies to tropomyosin isoforms in ulcerative colitis. *Gastroenterology* 1998;114:912-22.
25. Das KM, Sakamaki S, Vecchi M, et al. The production and characterization of monoclonal antibodies to a human colonic antigen associated with ulcerative colitis: cellular localization of the antigen by using the monoclonal antibody. *J Immunol* 1987;139:77-84.
26. Das KM, Vecchi M, Sakamaki S. A shared and unique epitope(s) on human colon, skin, and biliary epithelium detected by a monoclonal antibody. *Gastroenterology* 1990;98:464-9.
27. Bhagat S, Das KM. A shared and unique peptide in the human colon, eye, and joint detected by a monoclonal antibody. *Gastroenterology* 1994;107:103-8.
28. Parkes M, Cortes A, van Heel DA, et al. Genetic insights into common pathways and complex relationships among immune-mediated diseases. *Nat Rev Genet* 2013;14:661-73.
29. Severs M, van Erp SJ, van der Valk ME, et al. Smoking is Associated With Extra-intestinal Manifestations in Inflammatory Bowel Disease. *J Crohns Colitis* 2016;10:455-61.
30. Park JH, Peyrin-Biroulet L, Eisenhut M, et al. IBD immunopathogenesis: A comprehensive review of inflammatory molecules. *Autoimmun Rev* 2017;16:416-426.
31. Pastor Rojo O, Lopez San Roman A, Albeniz Arbizu E, et al. Serum lipopolysaccharide-binding protein in endotoxemic patients with inflammatory bowel disease. *Inflamm Bowel Dis* 2007;13:269-77.
32. Adenis A, Colombel JF, Lecouffe P, et al. Increased pulmonary and intestinal permeability in Crohn's disease. *Gut* 1992;33:678-82.
33. Sherlock JP, Joyce-Shaikh B, Turner SP, et al. IL-23 induces spondyloarthropathy by acting on ROR-gammat+ CD3+CD4-CD8- enthesal resident T cells. *Nat Med* 2012;18:1069-76.
34. Condliffe AM, Kitchen E, Chilvers ER. Neutrophil priming: pathophysiological consequences and underlying mechanisms. *Clin Sci (Lond)* 1998;94:461-71.
35. McCarthy DA, Rampton DS, Liu YC. Peripheral blood neutrophils in inflammatory bowel disease: morphological evidence of in vivo activation in active disease. *Clin Exp Immunol* 1991;86:489-93.
36. Nikolaus S, Bauditz J, Gionchetti P, et al. Increased secretion of pro-inflammatory cytokines by circulating polymorphonuclear neutrophils and regulation by interleukin 10 during intestinal inflammation. *Gut* 1998;42:470-6.
37. Smith AM, Rahman FZ, Hayee B, et al. Disordered macrophage cytokine secretion underlies impaired acute inflammation and bacterial clearance in Crohn's disease. *J Exp Med* 2009;206:1883-97.
38. Sanders TJ, McCarthy NE, Giles EM, et al. Increased production of retinoic acid by intestinal macrophages contributes to their inflammatory phenotype in patients with Crohn's disease. *Gastroenterology* 2014;146:1278-88 e1-2.
39. Clarke TB, Davis KM, Lysenko ES, et al. Recognition of peptidoglycan from the microbiota by Nod1 enhances systemic innate immunity. *Nat Med* 2010;16:228-31.
40. Balmer ML, Schurch CM, Saito Y, et al. Microbiota-derived compounds drive steady-state granulopoiesis via MyD88/TICAM signaling. *J Immunol* 2014;193:5273-83.
41. Iwamura C, Bouladoux N, Belkaid Y, et al. Sensing of the microbiota by NOD1 in mesenchymal stromal cells regulates murine hematopoiesis. *Blood* 2017;129:171-176.
42. Khosravi A, Yanez A, Price JG, et al. Gut microbiota promote hematopoiesis to control bacterial infection. *Cell Host Microbe* 2014;15:374-81.
43. Shi C, Jia T, Mendez-Ferrer S, et al. Bone marrow mesenchymal stem and progenitor cells induce monocyte emigration in response to circulating toll-like receptor ligands. *Immunity* 2011;34:590-601.
44. Griseri T, McKenzie BS, Schiering C, et al. Dysregulated hematopoietic stem and progenitor cell activity promotes interleukin-23-driven chronic intestinal inflammation. *Immunity* 2012;37:1116-29.
45. Askenase MH, Han SJ, Byrd AL, et al. Bone-Marrow-Resident NK Cells Prime Monocytes for Regulatory Function during Infection. *Immunity* 2015;42:1130-42.
46. Trompette A, Gollwitzer ES, Yadava K, et al. Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis. *Nat Med* 2014;20:159-66.
47. Kim YG, Udayanga KG, Totsuka N, et al. Gut dysbiosis promotes M2 macrophage polarization and allergic airway inflammation via fungi-induced PGE(2). *Cell Host Microbe* 2014;15:95-102.
48. Breban M, Tap J, Leboime A, et al. Faecal microbiota study reveals specific dysbiosis in spondyloarthritis. *Ann Rheum Dis* 2017;76:1614-1622.
49. Tito RY, Cypers H, Joossens M, et al. Brief Report: Dialister as a Microbial Marker of Disease Activity in Spondyloarthritis. *Arthritis Rheumatol* 2017;69:114-121.

50. Scher JU, Ubeda C, Artacho A, et al. Decreased bacterial diversity characterizes the altered gut microbiota in patients with psoriatic arthritis, resembling dysbiosis in inflammatory bowel disease. *Arthritis Rheumatol* 2015;67:128-39.
51. Eppinga H, Thio HB, Schreurs MWJ, et al. Depletion of *Saccharomyces cerevisiae* in psoriasis patients, restored by Dimethylfumarate therapy (DMF). *PLoS One* 2017;12:e0176955.
52. Sabino J, Vieira-Silva S, Machiels K, et al. Primary sclerosing cholangitis is characterised by intestinal dysbiosis independent from IBD. *Gut* 2016;65:1681-9.
53. Bajer L, Kverka M, Kostovcik M, et al. Distinct gut microbiota profiles in patients with primary sclerosing cholangitis and ulcerative colitis. *World J Gastroenterol* 2017;23:4548-4558.
54. Zinkernagel MS, Zysset-Burri DC, Keller I, et al. Association of the Intestinal Microbiome with the Development of Neovascular Age-Related Macular Degeneration. *Sci Rep* 2017;7:40826.
55. Wen X, Hu X, Miao L, et al. Epigenetics, microbiota, and intraocular inflammation: New paradigms of immune regulation in the eye. *Prog Retin Eye Res* 2018.
56. Crost EH, Tailford LE, Monestier M, et al. The mucin-degradation strategy of *Ruminococcus gnavus*: The importance of intramolecular trans-sialidases. *Gut Microbes* 2016;7:302-312.
57. Swann JR, Want EJ, Geier FM, et al. Systemic gut microbial modulation of bile acid metabolism in host tissue compartments. *Proc Natl Acad Sci U S A* 2011;108 Suppl 1:4523-30.
58. Hedin CR, van der Gast CJ, Stagg AJ, et al. The gut microbiota of siblings offers insights into microbial pathogenesis of inflammatory bowel disease. *Gut Microbes* 2017;8:359-365.
59. Hansen CH, Nielsen DS, Kverka M, et al. Patterns of early gut colonization shape future immune responses of the host. *PLoS One* 2012;7:e34043.
60. Montoya J, Matta NB, Suchon P, et al. Patients with ankylosing spondylitis have been breast fed less often than healthy controls: a case-control retrospective study. *Ann Rheum Dis* 2016;75:879-82.
61. van Sommeren S, Janse M, Karjalainen J, et al. Extraintestinal manifestations and complications in inflammatory bowel disease: from shared genetics to shared biological pathways. *Inflamm Bowel Dis* 2014;20:987-94.
62. Satsangi J, Grootcholten C, Holt H, et al. Clinical patterns of familial inflammatory bowel disease. *Gut* 1996;38:738-41.
63. Taleban S, Li D, Targan SR, et al. Ocular Manifestations in Inflammatory Bowel Disease Are Associated with Other Extra-intestinal Manifestations, Gender, and Genes Implicated in Other Immune-related Traits. *J Crohns Colitis* 2016;10:43-9.
64. Peeters H, Vander Cruyssen B, Laukens D, et al. Radiological sacroiliitis, a hallmark of spondylitis, is linked with CARD15 gene polymorphisms in patients with Crohn's disease. *Ann Rheum Dis* 2004;63:1131-4.
65. Martin TM, Smith JR, Rosenbaum JT. Anterior uveitis: current concepts of pathogenesis and interactions with the spondyloarthropathies. *Curr Opin Rheumatol* 2002;14:337-41.
66. Kontoyiannis D, Pasparakis M, Pizarro TT, et al. Impaired on/off regulation of TNF biosynthesis in mice lacking TNF AU-rich elements: implications for joint and gut-associated immunopathologies. *Immunity* 1999;10:387-98.
67. Bamias G, Dahman MI, Arseneau KO, et al. Intestinal-specific TNF α overexpression induces Crohn's-like ileitis in mice. *PLoS One* 2013;8:e72594.
68. Roulis M, Bongers G, Armaka M, et al. Host and microbiota interactions are critical for development of murine Crohn's-like ileitis. *Mucosal Immunol* 2016;9:787-97.
69. Apostolaki M, Manoloukos M, Roulis M, et al. Role of beta7 integrin and the chemokine/chemokine receptor pair CCL25/CCR9 in modeled TNF-dependent Crohn's disease. *Gastroenterology* 2008;134:2025-35.
70. Hammer RE, Maika SD, Richardson JA, et al. Spontaneous inflammatory disease in transgenic rats expressing HLA-B27 and human beta 2m: an animal model of HLA-B27-associated human disorders. *Cell* 1990;63:1099-112.
71. DeLay ML, Turner MJ, Klenk EI, et al. HLA-B27 misfolding and the unfolded protein response augment interleukin-23 production and are associated with Th17 activation in transgenic rats. *Arthritis Rheum* 2009;60:2633-43.
72. Turner MJ, Sowders DP, DeLay ML, et al. HLA-B27 misfolding in transgenic rats is associated with activation of the unfolded protein response. *J Immunol* 2005;175:2438-48.
73. Rath HC, Herfarth HH, Ikeda JS, et al. Normal luminal bacteria, especially *Bacteroides* species, mediate chronic colitis, gastritis, and arthritis in HLA-B27/human beta2 microglobulin transgenic rats. *J Clin Invest* 1996;98:945-53.

74. Ruutu M, Thomas G, Steck R, et al. beta-glucan triggers spondylarthritis and Crohn's disease-like ileitis in SKG mice. *Arthritis Rheum* 2012;64:2211-22.
75. Benham H, Rehaume LM, Hasnain SZ, et al. Interleukin-23 mediates the intestinal response to microbial beta-1,3-glucan and the development of spondyloarthritis pathology in SKG mice. *Arthritis Rheumatol* 2014;66:1755-67.
76. Milia AF, Ibba-Manneschi L, Manetti M, et al. HLA-B27 transgenic rat: an animal model mimicking gut and joint involvement in human spondyloarthritis. *Ann N Y Acad Sci* 2009;1173:570-4.
77. Fiorino G, Allez M, Malesci A, et al. Review article: anti TNF-alpha induced psoriasis in patients with inflammatory bowel disease. *Aliment Pharmacol Ther* 2009;29:921-7.
78. Fiorino G, Danese S, Pariente B, et al. Paradoxical immune-mediated inflammation in inflammatory bowel disease patients receiving anti-TNF- α agents. *Autoimmun Rev* 2014;13:15-9.
79. Palucka AK, Blanck JP, Bennett L, et al. Cross-regulation of TNF and IFN-alpha in autoimmune diseases. *Proc Natl Acad Sci U S A* 2005;102:3372-7.
80. de Gannes GC, Ghoreishi M, Pope J, et al. Psoriasis and pustular dermatitis triggered by TNF- α inhibitors in patients with rheumatologic conditions. *Arch Dermatol* 2007;143:223-31.
81. Nestle FO, Gilliet M. Defining upstream elements of psoriasis pathogenesis: an emerging role for interferon alpha. *J Invest Dermatol* 2005;125:xiv-xv.
82. Greuter T, Navarini A, Vavricka SR. Skin Manifestations of Inflammatory Bowel Disease. *Clin Rev Allergy Immunol* 2017.
83. Rahier JF, Buche S, Peyrin-Biroulet L, et al. Severe skin lesions cause patients with inflammatory bowel disease to discontinue anti-tumor necrosis factor therapy. *Clin Gastroenterol Hepatol* 2010;8:1048-55.
84. Vavricka SR, Galvan JA, Dawson H, et al. Expression Patterns of TNF α , MAdCAM1 and STAT3 in Intestinal and Skin Manifestations of Inflammatory Bowel Disease. *J Crohns Colitis* 2017.
85. Varkas G, Thevissen K, De Brabanter G, et al. An induction or flare of arthritis and/or sacroiliitis by vedolizumab in inflammatory bowel disease: a case series. *Ann Rheum Dis* 2017;76:878-881.
86. Tadbiri S, Peyrin-Biroulet L, Serrero M, et al. Impact of vedolizumab therapy on extra-intestinal manifestations in patients with inflammatory bowel disease: a multicentre cohort study nested in the OBSERV-IBD cohort. *Aliment Pharmacol Ther* 2018;47:485-493.
87. Hadis U, Wahl B, Schulz O, et al. Intestinal tolerance requires gut homing and expansion of FoxP3+ regulatory T cells in the lamina propria. *Immunity* 2011;34:237-46.
88. Cassani B, Villablanca EJ, Quintana FJ, et al. Gut-tropic T cells that express integrin $\alpha 4\beta 7$ and CCR9 are required for induction of oral immune tolerance in mice. *Gastroenterology* 2011;141:2109-18.
89. Tillack C, Ehmann LM, Friedrich M, et al. Anti-TNF antibody-induced psoriasiform skin lesions in patients with inflammatory bowel disease are characterised by interferon- γ -expressing Th1 cells and IL-17A/IL-22-expressing Th17 cells and respond to anti-IL-12/IL-23 antibody treatment. *Gut* 2014;63:567-77.
90. Andrisani G, Marzo M, Celleno L, et al. Development of psoriasis scalp with alopecia during treatment of Crohn's disease with infliximab and rapid response to both diseases to ustekinumab. *Eur Rev Med Pharmacol Sci* 2013;17:2831-6.
91. Gregoriou S, Kazakos C, Christofidou E, et al. Pustular psoriasis development after initial ustekinumab administration in chronic plaque psoriasis. *Eur J Dermatol* 2011;21:104-5.
92. Hueber W, Sands BE, Lewitzky S, et al. Secukinumab, a human anti-IL-17A monoclonal antibody, for moderate to severe Crohn's disease: unexpected results of a randomised, double-blind placebo-controlled trial. *Gut* 2012;61:1693-700.
93. Hohenberger M, Cardwell LA, Oussedik E, et al. Interleukin-17 inhibition: role in psoriasis and inflammatory bowel disease. *J Dermatolog Treat* 2017;1-6.
94. Brooklyn TN, Dunnill MG, Shetty A, et al. Infliximab for the treatment of pyoderma gangrenosum: a randomised, double blind, placebo controlled trial. *Gut* 2006;55:505-9.
95. Karreman MC, Luime JJ, Hazes JMW, et al. The Prevalence and Incidence of Axial and Peripheral Spondyloarthritis in Inflammatory Bowel Disease: A Systematic Review and Meta-analysis. *J Crohns Colitis* 2017;11:631-642.
96. Dougados M, Baeten D. Spondyloarthritis. *Lancet* 2011;377:2127-37.
97. Queiro R, Maiz O, Intxausti J, et al. Subclinical sacroiliitis in inflammatory bowel disease: a clinical and follow-up study. *Clin Rheumatol* 2000;19:445-9.
98. Peeters H, Vander Cruyssen B, Mielants H, et al. Clinical and genetic factors associated with sacroiliitis in Crohn's disease. *J Gastroenterol Hepatol* 2008;23:132-7.

99. de Vlam K, Mielants H, Cuvelier C, et al. Spondyloarthropathy is underestimated in inflammatory bowel disease: prevalence and HLA association. *J Rheumatol* 2000;27:2860-5.
100. Salvarani C, Vlachonikolis IG, van der Heijde DM, et al. Musculoskeletal manifestations in a population-based cohort of inflammatory bowel disease patients. *Scand J Gastroenterol* 2001;36:1307-13.
101. Bourikas LA, Papadakis KA. Musculoskeletal manifestations of inflammatory bowel disease. *Inflamm Bowel Dis* 2009;15:1915-24.
102. Rudwaleit M, van der Heijde D, Landewe R, et al. The development of Assessment of SpondyloArthritis international Society classification criteria for axial spondyloarthritis (part II): validation and final selection. *Ann Rheum Dis* 2009;68:777-83.
103. Rudwaleit M, van der Heijde D, Landewe R, et al. The Assessment of SpondyloArthritis International Society classification criteria for peripheral spondyloarthritis and for spondyloarthritis in general. *Ann Rheum Dis* 2011;70:25-31.
104. Sepriano A, Rubio R, Ramiro S, et al. Performance of the ASAS classification criteria for axial and peripheral spondyloarthritis: a systematic literature review and meta-analysis. *Ann Rheum Dis* 2017;76:886-890.
105. Belousova EA, D. Odintsova, A. Zakirov, R. Nagornykh, B. Protopopov, M. . Performance of ASAS criteria for inflammatory back pain in patients with inflammatory bowel disease. *Journal of Crohn's and Colitis* 2017;11.
106. Amor B, Dougados M, Mikiyawa M. [Criteria of the classification of spondylarthropathies]. *Rev Rhum Mal Osteoartic* 1990;57:85-9.
107. Dougados M, van der Linden S, Juhlin R, et al. The European Spondylarthropathy Study Group preliminary criteria for the classification of spondylarthropathy. *Arthritis Rheum* 1991;34:1218-27.
108. van der Linden S, Valkenburg HA, Cats A. Evaluation of diagnostic criteria for ankylosing spondylitis. A proposal for modification of the New York criteria. *Arthritis Rheum* 1984;27:361-8.
109. van der Heijde D, Lie E, Kvien TK, et al. ASDAS, a highly discriminatory ASAS-endorsed disease activity score in patients with ankylosing spondylitis. *Ann Rheum Dis* 2009;68:1811-8.
110. Mease P, Sieper J, Van den Bosch F, et al. Randomized controlled trial of adalimumab in patients with nonpsoriatic peripheral spondyloarthritis. *Arthritis Rheumatol* 2015;67:914-23.
111. Trusko B, Thorne J, Jabs D, et al. The Standardization of Uveitis Nomenclature (SUN) Project. Development of a clinical evidence base utilizing informatics tools and techniques. *Methods Inf Med* 2013;52:259-65, S1-6.
112. Jabs DA, Nussenblatt RB, Rosenbaum JT, et al. Standardization of uveitis nomenclature for reporting clinical data. Results of the First International Workshop. *Am J Ophthalmol* 2005;140:509-16.
113. Vavricka SR, Brun L, Ballabeni P, et al. Frequency and risk factors for extraintestinal manifestations in the Swiss inflammatory bowel disease cohort. *Am J Gastroenterol* 2011;106:110-9.
114. Vavricka SR, Schoepfer A, Scharl M, et al. Extraintestinal Manifestations of Inflammatory Bowel Disease. *Inflamm Bowel Dis* 2015;21:1982-92.
115. Fredriksson T, Pettersson U. Severe psoriasis--oral therapy with a new retinoid. *Dermatologica* 1978;157:238-44.
116. Marks R, Barton SP, Shuttleworth D, et al. Assessment of disease progress in psoriasis. *Arch Dermatol* 1989;125:235-40.
117. Ramsay B, Lawrence CM. Measurement of involved surface area in patients with psoriasis. *Br J Dermatol* 1991;124:565-70.
118. Hanifin JM, Thurston M, Omoto M, et al. The eczema area and severity index (EASI): assessment of reliability in atopic dermatitis. EASI Evaluator Group. *Exp Dermatol* 2001;10:11-8.
119. Weizman A, Huang B, Berel D, et al. Clinical, serologic, and genetic factors associated with pyoderma gangrenosum and erythema nodosum in inflammatory bowel disease patients. *Inflamm Bowel Dis* 2014;20:525-33.
120. Alonso A, Domenech E, Julia A, et al. Identification of risk loci for Crohn's disease phenotypes using a genome-wide association study. *Gastroenterology* 2015;148:794-805.
121. International Genetics of Ankylosing Spondylitis C, Cortes A, Hadler J, et al. Identification of multiple risk variants for ankylosing spondylitis through high-density genotyping of immune-related loci. *Nat Genet* 2013;45:730-8.
122. Reveille JD. Biomarkers for diagnosis, monitoring of progression, and treatment responses in ankylosing spondylitis and axial spondyloarthritis. *Clin Rheumatol* 2015;34:1009-18.
123. Prajzlerova K, Grobelna K, Pavelka K, et al. An update on biomarkers in axial spondyloarthritis. *Autoimmun Rev* 2016;15:501-9.

124. Menti E, Lanera C, Lorenzoni G, et al. Bayesian Machine Learning Techniques for revealing complex interactions among genetic and clinical factors in association with extra-intestinal Manifestations in IBD patients. *AMIA Annu Symp Proc* 2016;2016:884-893.
125. van Gaalen FA, Bakker PA, de Hooze M, et al. Assessment of sacroiliitis by radiographs and MRI: where are we now? *Curr Opin Rheumatol* 2014;26:384-8.
126. Atzeni F, Defendenti C, Ditto MC, et al. Rheumatic manifestations in inflammatory bowel disease. *Autoimmun Rev* 2014;13:20-3.
127. Hermann KG, Baraliakos X, van der Heijde DM, et al. Descriptions of spinal MRI lesions and definition of a positive MRI of the spine in axial spondyloarthritis: a consensual approach by the ASAS/OMERACT MRI study group. *Ann Rheum Dis* 2012;71:1278-88.
128. Sudol-Szopinska I, Pracon G. Diagnostic imaging of psoriatic arthritis. Part II: magnetic resonance imaging and ultrasonography. *J Ultrason* 2016;16:163-74.
129. D'Agostino MA, Aegerter P, Bechara K, et al. How to diagnose spondyloarthritis early? Accuracy of peripheral enthesitis detection by power Doppler ultrasonography. *Ann Rheum Dis* 2011;70:1433-40.
130. Maksymowych WP. Progress in spondylarthritis. Spondyloarthritis: lessons from imaging. *Arthritis Res Ther* 2009;11:222.
131. van der Heijde D, Landewe R, van der Linden S. How should treatment effect on spinal radiographic progression in patients with ankylosing spondylitis be measured? *Arthritis Rheum* 2005;52:1979-85.
132. Spoorenberg A, de Vlam K, van der Linden S, et al. Radiological scoring methods in ankylosing spondylitis. Reliability and change over 1 and 2 years. *J Rheumatol* 2004;31:125-32.
133. Baraliakos X, Landewe R, Hermann KG, et al. Inflammation in ankylosing spondylitis: a systematic description of the extent and frequency of acute spinal changes using magnetic resonance imaging. *Ann Rheum Dis* 2005;64:730-4.
134. Braun J, Baraliakos X, Golder W, et al. Magnetic resonance imaging examinations of the spine in patients with ankylosing spondylitis, before and after successful therapy with infliximab: evaluation of a new scoring system. *Arthritis Rheum* 2003;48:1126-36.
135. MacKay K, Mack C, Brophy S, et al. The Bath Ankylosing Spondylitis Radiology Index (BASRI): a new, validated approach to disease assessment. *Arthritis Rheum* 1998;41:2263-70.
136. Van Praet L, Van den Bosch FE, Jacques P, et al. Microscopic gut inflammation in axial spondyloarthritis: a multiparametric predictive model. *Ann Rheum Dis* 2013;72:414-7.
137. Mundwiler ML, Mei L, Landers CJ, et al. Inflammatory bowel disease serologies in ankylosing spondylitis patients: a pilot study. *Arthritis Res Ther* 2009;11:R177.
138. Wallis D, Asaduzzaman A, Weisman M, et al. Elevated serum anti-flagellin antibodies implicate subclinical bowel inflammation in ankylosing spondylitis: an observational study. *Arthritis Res Ther* 2013;15:R166.
139. Jiang L, Xia B, Li J, et al. Retrospective survey of 452 patients with inflammatory bowel disease in Wuhan city, central China. *Inflamm Bowel Dis* 2006;12:212-7.
140. Burisch J, Pedersen N, Cukovic-Cavka S, et al. East-West gradient in the incidence of inflammatory bowel disease in Europe: the ECCO-EpiCom inception cohort. *Gut* 2014;63:588-97.
141. Ng SC, Zeng Z, Niewiadomski O, et al. Early Course of Inflammatory Bowel Disease in a Population-Based Inception Cohort Study From 8 Countries in Asia and Australia. *Gastroenterology* 2016;150:86-95 e3; quiz e13-4.
142. Lakatos L, Pandur T, David G, et al. Association of extraintestinal manifestations of inflammatory bowel disease in a province of western Hungary with disease phenotype: results of a 25-year follow-up study. *World J Gastroenterol* 2003;9:2300-7.
143. Roberts H, Rai SN, Pan J, et al. Extraintestinal manifestations of inflammatory bowel disease and the influence of smoking. *Digestion* 2014;90:122-9.
144. Karmiris K, Avgerinos A, Tavernaraki A, et al. Prevalence and Characteristics of Extra-intestinal Manifestations in a Large Cohort of Greek Patients with Inflammatory Bowel Disease. *J Crohns Colitis* 2016;10:429-36.
145. Orchard TR, Wordsworth BP, Jewell DP. Peripheral arthropathies in inflammatory bowel disease: their articular distribution and natural history. *Gut* 1998;42:387-91.
146. Vegh Z, Kurti Z, Gonczi L, et al. Association of extraintestinal manifestations and anaemia with disease outcomes in patients with inflammatory bowel disease. *Scand J Gastroenterol* 2016;51:848-54.
147. Isene R, Bernklev T, Hoie O, et al. Extraintestinal manifestations in Crohn's disease and ulcerative colitis: results from a prospective, population-based European inception cohort. *Scand J Gastroenterol* 2015;50:300-5.

148. Ott C, Takses A, Obermeier F, et al. Smoking increases the risk of extraintestinal manifestations in Crohn's disease. *World J Gastroenterol* 2014;20:12269-76.
149. Bernstein CN, Blanchard JF, Rawsthorne P, et al. The prevalence of extraintestinal diseases in inflammatory bowel disease: a population-based study. *Am J Gastroenterol* 2001;96:1116-22.
150. Manguso F, Sanges M, Staiano T, et al. Cigarette smoking and appendectomy are risk factors for extraintestinal manifestations in ulcerative colitis. *Am J Gastroenterol* 2004;99:327-34.
151. Orchard TR, Holt H, Bradbury L, et al. The prevalence, clinical features and association of HLA-B27 in sacroiliitis associated with established Crohn's disease. *Aliment Pharmacol Ther* 2009;29:193-7.
152. Orchard TR, Chua CN, Ahmad T, et al. Uveitis and erythema nodosum in inflammatory bowel disease: clinical features and the role of HLA genes. *Gastroenterology* 2002;123:714-8.
153. Orchard TR, Thiyagaraja S, Welsh KI, et al. Clinical phenotype is related to HLA genotype in the peripheral arthropathies of inflammatory bowel disease. *Gastroenterology* 2000;118:274-8.
154. Klingberg E, Strid H, Stahl A, et al. A longitudinal study of fecal calprotectin and the development of inflammatory bowel disease in ankylosing spondylitis. *Arthritis Res Ther* 2017;19:21.
155. Peyrin-Biroulet L, Van Assche G, Gomez-Ulloa D, et al. Systematic Review of Tumor Necrosis Factor Antagonists in Extraintestinal Manifestations in Inflammatory Bowel Disease. *Clin Gastroenterol Hepatol* 2017;15:25-36 e27.
156. Hirten R, Longman RS, Bosworth BP, et al. Vedolizumab and Infliximab Combination Therapy in the Treatment of Crohn's Disease. *Am J Gastroenterol* 2015;110:1737-8.
157. Vande Casteele N, Herfarth H, Katz J, et al. American Gastroenterological Association Institute Technical Review on the Role of Therapeutic Drug Monitoring in the Management of Inflammatory Bowel Diseases. *Gastroenterology* 2017;153:835-857 e6.
158. Peyrin-Biroulet L, Panes J, Sandborn WJ, et al. Defining Disease Severity in Inflammatory Bowel Diseases: Current and Future Directions. *Clin Gastroenterol Hepatol* 2016;14:348-354 e17.
159. Pincus T, Gibofsky A, Weinblatt ME. Urgent care and tight control of rheumatoid arthritis as in diabetes and hypertension: better treatments but a shortage of rheumatologists. *Arthritis Rheum* 2002;46:851-4.
160. Rachmani R, Slavacheski I, Berla M, et al. Treatment of high-risk patients with diabetes: motivation and teaching intervention: a randomized, prospective 8-year follow-up study. *J Am Soc Nephrol* 2005;16 Suppl 1:S22-6.
161. Eeg-Olofsson K, Cederholm J, Nilsson PM, et al. Glycemic and risk factor control in type 1 diabetes: results from 13,612 patients in a national diabetes register. *Diabetes Care* 2007;30:496-502.
162. Patel A, Group AC, MacMahon S, et al. Effects of a fixed combination of perindopril and indapamide on macrovascular and microvascular outcomes in patients with type 2 diabetes mellitus (the ADVANCE trial): a randomised controlled trial. *Lancet* 2007;370:829-40.
163. Peyrin-Biroulet L, Sandborn W, Sands BE, et al. Selecting Therapeutic Targets in Inflammatory Bowel Disease (STRIDE): Determining Therapeutic Goals for Treat-to-Target. *Am J Gastroenterol* 2015;110:1324-38.
164. FDA. Guidance for Industry: Patient-Reported Outcome Measures: Use in Medical Product Development to Support Labeling Claims. In: Office of Communications DoDI, Center for Drug Evaluation and Research, Food and Drug Administration, ed, 2009.
165. Guyatt G, Mitchell A, Irvine EJ, et al. A new measure of health status for clinical trials in inflammatory bowel disease. *Gastroenterology* 1989;96:804-10.
166. Love JR, Irvine EJ, Fedorak RN. Quality of life in inflammatory bowel disease. *J Clin Gastroenterol* 1992;14:15-9.
167. Reilly MC, Zbrozek AS, Dukes EM. The validity and reproducibility of a work productivity and activity impairment instrument. *Pharmacoeconomics* 1993;4:353-65.
168. Ware J, Jr., Kosinski M, Keller SD. A 12-Item Short-Form Health Survey: construction of scales and preliminary tests of reliability and validity. *Med Care* 1996;34:220-33.
169. Cheung KO, M. Oppe, M. Rabin, R. EQ-5D User Guide: Basic Information on how to use the EQ-5D Version 2.0. Rotterdam: EuroQoL Group, 2009.